

ASYMMETRIC SYNTHESIS OF CHI-CONSTRAINED GLUTAMIC ACIDS AND RELATED COMPOUNDS VIA MICHAEL ADDITION REACTIONS

Alicja Wzorek¹, Alexander E. Sorochinsky², Karel D. Klika³,
Taizo Ono⁴, Jianlin Han^{5*}, Vadim A. Soloshonok^{6,7*}

¹ Institute of Chemistry, Jan Kochanowski University in Kielce, Uniwersytecka 7, 25–406 Kielce, Poland;

² Department of Fine Organic Synthesis, V.P. Kukhar Institute of Bioorganic Chemistry and Petrochemistry, The National Academy of Sciences of Ukraine, 1 Murmanska str., Kyiv 02094, Ukraine;

³ Molecular Structure Analysis, German Cancer Research Center (DKFZ), ImNeuenheimer Feld 280, 69120 Heidelberg, Germany;

⁴ National Institute of Advanced Industrial Science and Technology, 463–8560, Nagoya, Japan;

⁵ Jiangsu Co-Innovation Center of Efficient Processing and Utilization of Forest Resources, College of Chemical Engineering, Nanjing Forestry University, Nanjing 210037, China;

⁶ Department of Organic Chemistry I, Faculty of Chemistry, University of the Basque Country UPV/EHU, Paseo Manuel Lardizábal 3, 20018 San Sebastián, Spain;

⁷ IKERBASQUE, Basque Foundation for Science, María Díaz de Haro 3, Plaza Bizkaia, 48013 Bilbao, Spain.

e-mail: vadimsoloshonok@gmail.com

Michael addition reactions involving nucleophilic glycine equivalents and α,β -unsaturated carboxylic acid derivatives offer a concise and generalized methodological approach to synthesizing a family of χ -constrained five-carbon-atom amino acids. These amino acids play a crucial role in de novo peptide design and the elucidation of peptide/protein three-dimensional structures and their biological functions/activities. This review encapsulates the significant synthetic and methodological advancements in the field to date. Each method discussed includes an evaluation of synthetic opportunities and limitations, practicality and efficiency of the procedures, and mechanistic rationale behind the observed stereochemical preferences.

Keywords: Asymmetric synthesis, Michael additions, glycine equivalents, metal complexes, Schiff bases, conformations, steric constrain. Glutamic acid, pyroglutamic acid, proline.

INTRODUCTION. Amino acids (AAs) are fundamental to the fabric of life, acting as the building blocks of proteins that drive countless biological processes [1]. In general sciences, their study has provided crucial insights into the mechanisms of life, from understanding metabolic pathways to uncovering the genetic code. Researchers delve into amino acid sequences to elucidate protein structure and function, enabling advances in fields like biochemistry, molecular biology, and genetics. The versatility and functionality of AAs make them indispensable for exploring the intricacies of cellular functions and organismal development [2–6].

In the pharmaceutical industry, amino acids hold a pivotal role in the development of drugs and therapeutic agents [7–10]. Many medications are designed to mimic or modulate the activity of naturally occurring AAs and their derivatives, targeting specific pathways to treat a variety of diseases. For instance, amino acid derivatives are used in the synthesis of antiviral drugs, antibiotics, and treatments for metabolic disorders [11–13]. Additionally, the role of AAs in protein synthesis is harnessed to develop biologics, such as monoclonal antibodies and vaccines, which are essential for modern medicine. The continuous exploration of AAs and their properties drives innovation, leading to more effective and targeted therapies that improve human health and well-being [14–20].

Glutamic acid and its cyclic derivative, pyroglutamic acid, play pivotal roles in peptides and drug design [21]. Glutamic acid, an essential amino acid, contributes to protein structure and function through its side chain, which can participate in ionic interactions and hydrogen bonding. This property is critical in stabilizing protein structures and facilitating enzyme ca-

talysis [22]. Pyroglutamic acid, formed by the cyclization of glutamic acid, introduces rigidity into peptide chains, thereby reducing conformational flexibility. This structural constraint is advantageous in drug design, as it enhances binding specificity and stability, improving the efficacy of peptide-based therapeutics [23]. Both glutamic and pyroglutamic acids are integral in designing peptides with optimized pharmacological properties, making them invaluable in the development of novel drugs and therapeutic agents [24].

In this review article, we will discuss the synthesis of a specialized class of glutamic acid and related compounds: stereochemically constrained glutamic/pyroglutamic acids formed via the Michael addition reaction of nucleophilic glycine equivalents. These amino acids (AAs) adopt a relatively limited number of conformations and play a crucial role in the *de novo* design of peptides with predetermined 3D structures and enhanced peptide-receptor interactions.

*Role of $\chi(chi)$ -constrained amino acids in the *de novo* design of peptides and peptidomimetics.*

Reducing the number of amino acid (AA) conformations is crucial for optimizing peptide-receptor interactions, which play a vital role in various biological processes and therapeutic applications [25]. By limiting the conformational flexibility of peptides, researchers can design more specific and stable interactions between peptides and their target receptors. This precision enhances binding affinity and selectivity, leading to improved efficacy and reduced off-target effects in drug development. Tailoring peptide structures to adopt fewer conformations not only streamlines the identification of potent therapeutic candi-

dates but also contributes to the stability and bioavailability of the peptides in physiological environments. Ultimately, this approach holds promise for developing highly effective and targeted treatments for a wide range of diseases [26].

The 3D structure of peptides is determined by three key components: peptide sequence (primary structure), conformation (secondary structure), and the topographical positioning of side-chain functional groups. While the importance of peptide sequence and conformation in biological activity has been widely acknowledged, the third factor – the spatial arrangement of side chains on the peptide backbone (referred to as χ -space) – has only recently gained attention [26]. The importance of the torsional angles ϕ (phi), ψ (psi), and ω (omega) (Figure 1. **A**) in determining the 3D structure of the peptide backbone [27–30], as well as the χ (chi) torsional angles (Figure 1, **B**) in defining the position of side-chain functional groups, for elucidating peptide biological activity, has been demonstrated [31–37]. While ϕ , ψ , and ω angles are generally influenced by the nature of amino acid residues and the peptide's secondary structure (e.g., cyclization or global constraints), controlling χ angles is much more challenging.

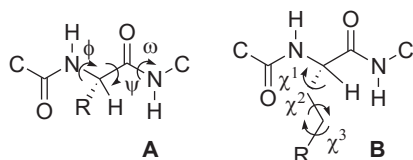


Fig. 1. Dihedral angles ϕ (phi), ψ (psi), ω (omega) (**A**), and χ (chi) (**B**), leading to numerous conformations of amino acid residues in peptides.

It was demonstrated that, in contrast to α -substitution in amino acids, which has little

effect on the χ angles, the introduction of a methyl group in the β -position substantially enhances the population of one out of three [gauche-(+), trans, and gauche-(-)] χ 1-rotamers. For instance, as illustrated for methyl-phenylalanines **1** (Figure 2), unfavorable steric interactions between the vicinal substituents (β -methyl and amino/carboxylic groups) resulted in a strong preference for the corresponding trans rotamer over the two gauche conformations [38, 39]. Considering β -substituted prolines **2**, as proline-phenylalanine chimeras, one could expect that rotational freedom in these derivatives might be much more limited for χ 1, compared to that of β -methyl substituted phenylalanine **1** [40–42]. Indeed, it was shown that in both cis and trans diastereomers of **2**, the corresponding (-)-gauche conformers are physically inaccessible. Moreover, molecular calculations of proline-tyrosine chimeras of type **2** revealed that the trans rotamer is preferred by up to 2 kcal/mol over the corresponding (-)-gauche conformer [43–64].

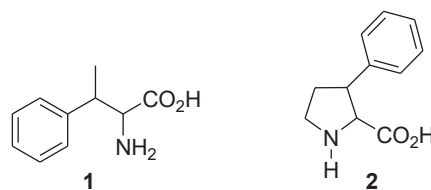


Fig. 2. β -Substitution and proline chimera with limited rotational freedom around the χ 1 (chi) torsional angle.

The challenge associated with the asymmetric synthesis of χ -constrained amino acids stems from their sterically congested nature, the presence of at least two stereogenic centers, and the necessity of preparing all (in most cases four) stereoisomers for SAR studies. The

unique potential of χ -constrained amino acids in the rational design of peptides and peptide mimetics has generated significant research efforts aimed at developing various synthetic methods to prepare these amino acids in stereochemically defined forms.

Asymmetric Michael addition reactions of glycine equivalents.

The ever-growing demand for proteinogenic and tailor-made amino acids has spurred extensive research in the field of asymmetric synthesis of amino acids [55–64]. Chiral glycine equivalents have garnered significant attention, as nucleophilic or electrophilic homologation of glycine offers a generalized approach to synthesizing various α -amino acids. The primary challenge in designing chiral nucleophilic glycine equivalents lies in controlling enolate face selectivity, i.e., the configuration of the α -stereogenic center in the resulting amino acids. Conversely, the Michael addition reactions of glycine equivalents with β -substituted acrylic acid derivatives present an issue with Michael acceptor face selectivity, crucial for determining the configuration of the β -carbon stereogenic center.

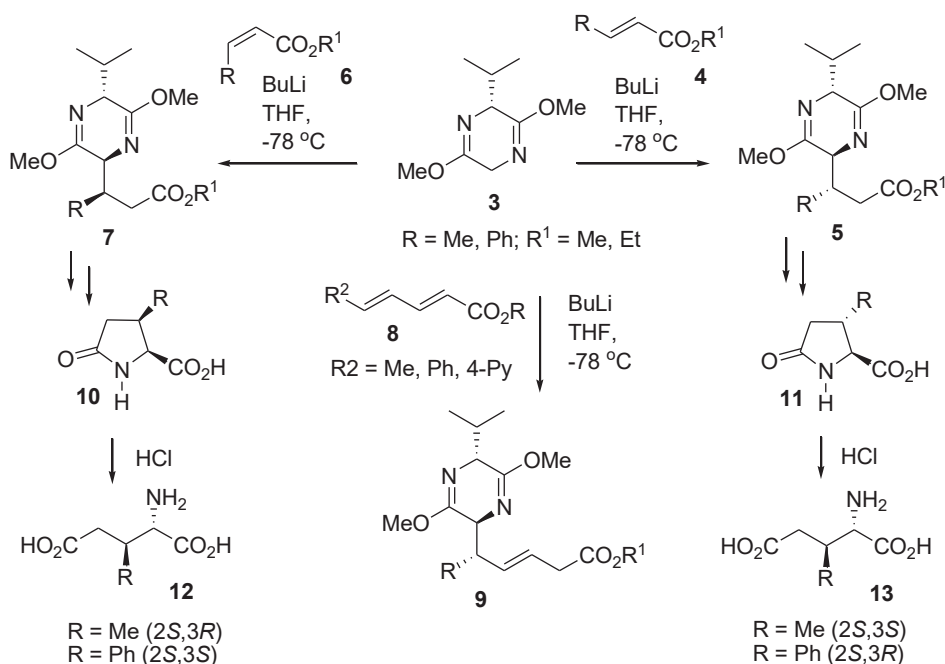
Most well-designed chiral glycine equivalents achieve high levels of stereocontrol at the α -stereogenic carbon when reacting with β -substituted acrylic acid derivatives, but they are less effective in controlling the absolute configuration at the β -position. However, in certain cases, the stereochemical requirements of both the chiral glycine derivative and the Michael acceptor can align, resulting in a synthetically valuable stereochemical outcome.

Bis-lactim ether methodology (Scheme 1) [65] has been widely used for preparing var-

ious α -amino acids. The corresponding anion is typically generated by treating the bis-lactim ether of cyclo-(R)-Val-Gly **3** with BuLi at $-78\text{ }^{\circ}\text{C}$. Electrophilic attack on the in situ generated anion by alkyl halides, carbonyl compounds, or acrylic acid derivatives occurs almost exclusively from the least hindered face, away from the bulky iso-Pr group, giving rise to the corresponding derivatives of α -amino acids with an α -(S) absolute configuration and very high enantiomeric excess (ee) [65].

In the original protocol [66], the Michael addition of alkyl crotonates **4** to (R)-**3** produced a mixture of two diastereomeric products, (2S, 3R) and (2S, 3S) **5**, in a ratio of 3 to 1. However, another report on the same reaction claimed much higher stereoselectivity, with no other diastereomers identified among the by-products [67]. The reactions of **3** with alkyl trans-cinnamates led to the formation of the corresponding derivatives of (2S, 3R)-3-phenylglutamic acid [68] in synthetically useful chemical yields (>75%) and with high diastereoselectivity (>90% de). In this case, there is no discrepancy between the original report [66] and the following reports reproducing these additions on a relatively large scale (>25 g) [69].

The *bis*-lactim ether method also provides access to cis-diastereomers of glutamic/pyroglutamic acids **10** as illustrated in Scheme 1, through additions of **3** with cis-configured Michael acceptors. Thus, the reactions of **3** with alkyl cis-crotonates and -cinnamates gave rise to the corresponding addition products **7** as the major product, along with three other diastereomeric products [65–67, 69]. The stereochemical outcome in this case is slightly lower compared to the additions with trans-configured Michael acceptors.

Scheme 1. *bis*-Lactim ether methodology for Machiel addition reactions.

The additions between (S)-3 and methyl 2,4-pentadienoates **8** (Scheme 1) were reported to proceed with virtually complete diastereoselectivity [70]. The reaction conducted under standard conditions for this method resulted in only one detectable diastereomer by NMR, with spectral characteristics consistent with the corresponding 1,6-addition product **9**. Unfortunately, the stereochemistry of the β -stereogenic center of the newly formed amino acid residue was not determined, while the configuration of the α -carbon atom was found to be (R) [70].

In principle, the *bis*-lactim ether method allows for generalized access to various β -alkyl and β -aryl substituted glutamic/pyroglutamic acids, including both *cis* and *trans*-diastereomeric forms. However, this method has several inherent drawbacks that make it less attractive from a synthetic standpoint. Be-

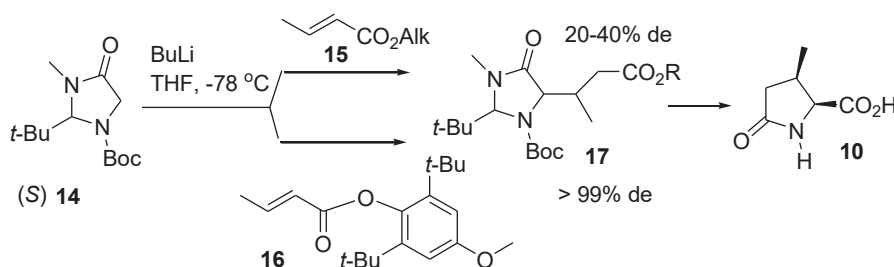
sides usually moderate chemical yields and, in most cases, incomplete stereoselectivity at the β -stereogenic carbon of the target glutamic/pyroglutamic acids, hydrolysis of the addition products **7**, **5**, **9** to the target amino acids involves painstaking separations, usually by distillation, of the resultant amino acid esters from methyl valinate coming from the starting chiral auxiliary. Since this type of separation depends on the physical properties of the compounds, it necessitates the development of a work-up procedure for each particular case.

Another highly diastereoselective approach to β -substituted glutamic/pyroglutamic acids is based on Michael addition reactions of the enolate of (S)-imidazolidinone **14** (Scheme 2). These reactions were studied using conventional alkyl esters (Me, Et) of β -substituted acrylic acids **15** as well as specially designed «sterically protected but electronically effective»

2,6-di-*t*-butyl-4-methoxyphenyl esters **16** [71, 72]. The stereochemical outcome of the additions between anion (*S*)-**14** and alkyl esters **15** was rather disappointing. The addition products **17** were isolated in moderate chemical yields [68% (Me), 78% (Et)] and with low diastereoselectivity (20–40% de).

In contrast, the reactions of (*S*)-**14** with 2,6-di-*t*-butyl-4-methoxyphenyl esters **16** oc-

curred at higher reaction rates, resulting in sole diastereomeric products **17** with good chemical yields (78–96%) and complete (>99% de) diastereoselectivity. The stereochemical outcome in these addition reactions is assumed to be kinetically controlled. It is important to emphasize that this approach offers *cis*-pyroglutamic acids, which are hardly attainable by the *bis*-lactim ether method.



Scheme 2. Imidazolidinone-derived chiral glycine equivalent in Michael addition reactions.

The mechanistic rationale for the very high stereoselectivity at the α -stereogenic center of the newly formed amino acids is rather straightforward, as the Michael acceptor attacks the corresponding anion generated from **14** almost exclusively from the side opposite the bulky *t*-butyl group. On the other hand, explaining the virtually complete face- and diastereoselectivity observed in the additions of (*S*)-**14** with bulky aryl esters **16** was not straightforward. Molecular mechanics calculations revealed that the energy difference between two conformers, *s*-*cis* **18** and *s*-*trans* **19** (Fig. 3), of the Michael acceptor is minimal. However, the calculations suggested that the *s*-*cis* **18** conformer should be substantially more reactive than the *s*-*trans* **19** because the reactive β -carbon atom in the former is shielded by bulky *t*-butyl groups. This rationale seems reasonable and, most importantly, emphasizes for the first time the relevance

of *s*-*cis* and *s*-*trans* conformers of the Michael acceptor to the stereochemical outcome in these addition reactions.

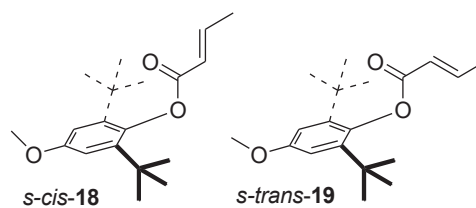


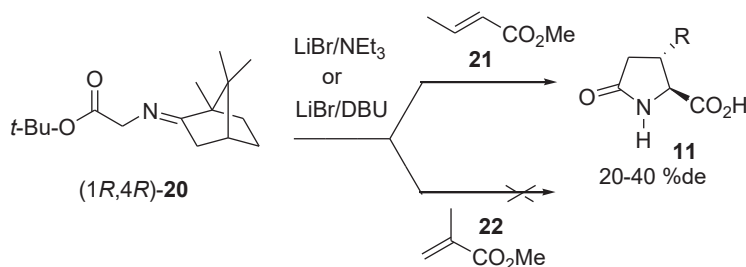
Fig. 3. *s*-*cis* and *s*-*trans* Conformers of 2,6-di-*t*-butyl-4-methoxyphenyl esters **16**.

Considering the substrate generality (alkyl, *i*-Pr, Bn, aryl groups), high chemical yields, and virtually complete diastereoselectivity, the imidazolidinone method could be regarded as nearly perfect for preparing *cis*-configured pyroglutamic/glutamic acids and their derivatives. However, from the standpoint of synthetic efficiency and practicality, this method is hardly at-

tractive for multigram preparation of the target compounds. Besides the general disadvantages of the Seebach method, such as the generation of the enolate of **14** at -78°C with BuLi and the relatively drastic reaction conditions required to remove the chiral auxiliary (0.75 N HCl for 16 h at 105°C), the multistage (3–4 stages) preparation of the aryl esters **16**, as well as the oxidative deprotection of the ester moiety with Ce(IV) [73–77], render the whole procedure problematic for large-scale preparations.

The application of chiral equivalents of nucleophilic glycine, characterized by the high CH acidity of the glycine methylene group, offers a significant synthetic advantage. In this approach, the corresponding enolate can be generated under mild reaction conditions,

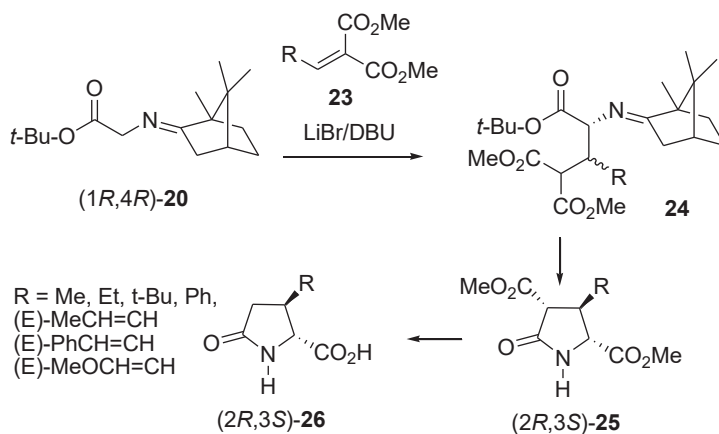
which are much easier to control and more practical than using highly reactive and hygroscopic BuLi at -78°C . It was demonstrated that the generation of the corresponding enolate from Schiff base **20** (Scheme 3) could be easily achieved by treating **20**, derived from t-butyl glycinate and natural (1R, 4R)-camphor, with a mixture of LiBr and triethylamine in THF [78]. The enolate thus generated readily reacted at room temperature with methyl acrylate **21**, furnishing a mixture of two b-(R)- and b-(S)-diastereomers in a ratio of up to 84/16, respectively, indicating relatively poor face-diastereoselectivity in the addition. The Michael addition of **20** with methyl crotonate **22** completely failed, presumably due to the greater steric bulkiness and lower electrophilicity of the latter.



Scheme 3. Michael addition reactions of Schiff base **20** derived from t-butyl glycinate and (1R, 4R)-camphor.

It was found that the application of α -methoxycarbonyl-substituted derivatives **23** (Scheme 4) led to a significant increase in the diastereoselectivity of the additions. Except for the reaction of **20** with dimethyl ethylidenemalonate **23** (R = Me), which furnished a mixture of two diastereomeric products (ratio 86/15), all other studied additions yielded only a single diastereomer in the reaction mixture. The remarkable difference in the stereochemical outcomes between the reactions of ethylidenemalonate **23** (R = Me) (72% de) and

propylidenemalonate **23** (R = Et) (over 99% de) with enolate of **20** was attributed by the authors to the simple difference in steric bulk between the Me and Et groups. To the best of our knowledge, this is a rare example of such dramatic stereochemical consequences resulting from the difference in steric bulk between methyl and ethyl groups [79]. The transformation of the addition products **24** to the final amino acids **26** involves simple hydrolytic removal of the chiral auxiliary and decarboxylation of the intermediate **25**.



Scheme 4. Michael addition reactions of Schiff base **20** with α -methoxycarbonyl-substituted acrylic acids **23**.

This method appears to be quite general for preparing various β -substituted pyroglutamic/glutamic acid derivatives. Of particular importance is the successful reaction of 2,2-dimethylpropylidenemalonate **23** ($\text{R} = \text{t-Bu}$) with **20**, which ultimately leads to sterically constrained β -t-butyl derivatives of the target amino acids **26**. Notably, in contrast to the *bis*-lactim ether methodology (Scheme 1), the reactions between **20** and derivatives **23** containing conjugated C,C double bonds featured only 1,4-addition. This allowed for the preparation of the corresponding pyroglutamic/glutamic acids bearing potentially useful unsaturated functionality. However, the general applicability of this method is greatly limited by the use of naturally occurring camphor as a chiral auxiliary, as its (1S, 4S) enantiomeric form is scarcely available.

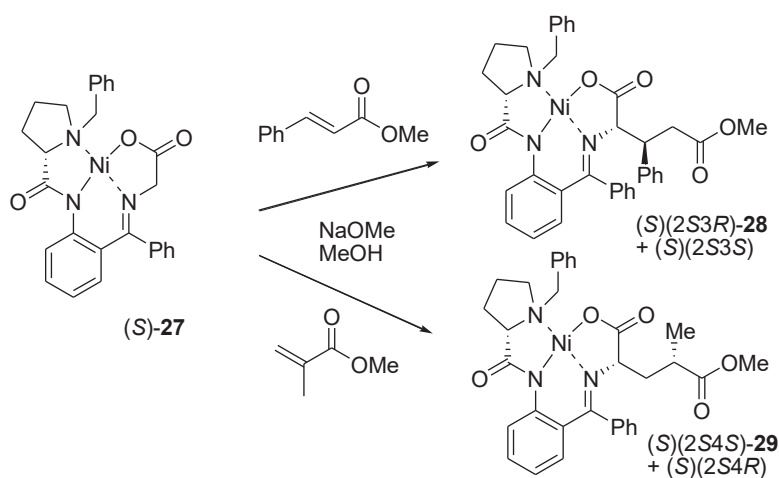
Another example of Michael addition reactions between chiral Schiff bases of glycine and α , β -unsaturated carboxylic acid derivatives was developed based on Ni(II) complex **27** shown in Scheme 5 [80–83]. The Ni(II) complex **27** was found to react easily in MeOH and in the presence of NaOMe as a base, with me-

thyl cinnamate and methacrylate, giving rise to the corresponding diastereomeric products **28** and **29**. In both cases, the enolate face-diastereoselectivity was very high as α -(R)-configured products were not detected in noticeable amounts. However, diastereoselectivity at C-3 and C-4 positions was disappointingly low, furnishing the products in approximately a 2/1 and 1/1 ratio respectively. It is interesting to note that in this case the stereochemical outcome is kinetically controlled, these reactions are highly reversible and thus the ratios of diastereomeric products are thermodynamically controlled.

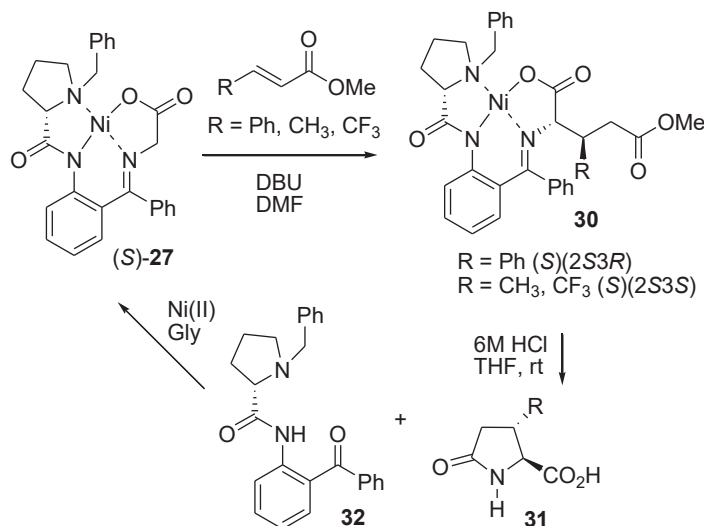
Taking advantage of high C-H acidity of the glycine methylene group in **27**, it was found that the corresponding Michael addition reactions with ethyl crotonate, 4,4,4-trifluorocrotonate and cinnamate [84,85] can be conducted in DMF in the presence of DBU as a base. The stereochemical outcome of the reactions was found to be kinetically controlled. The diastereoselectivity in the additions of (S)-**27** with ethyl crotonate and cinnamate was found to be noticeably better compared to the outcomes observed under the thermodynamically con-

trolled conditions (Scheme 5). The major products **30** ($R = \text{Me}, \text{Ph}$) were obtained in a ratio of 4/1, respectively. The stereochemical outcome of the reaction between (*S*)-**27** and ethyl 4,4,4-trifluorocrotonate was slightly higher, with a diastereomeric ratio of up to 5.6/1. This can be attributed to the more sterically demanding nature of the trifluoromethyl group

[86–88]. The major diastereomers **30** were purified by crystallization of the reaction mixture and disassembled to afford the target pyroglutamic acids **31**, along with the recovery of the chiral ligand (*S*)-**32**. The ability to recycle the chiral ligand (*S*)-**32**, which can be reused for preparation of new batches of Ni(II) complex **27** is a significant advantage of this method.



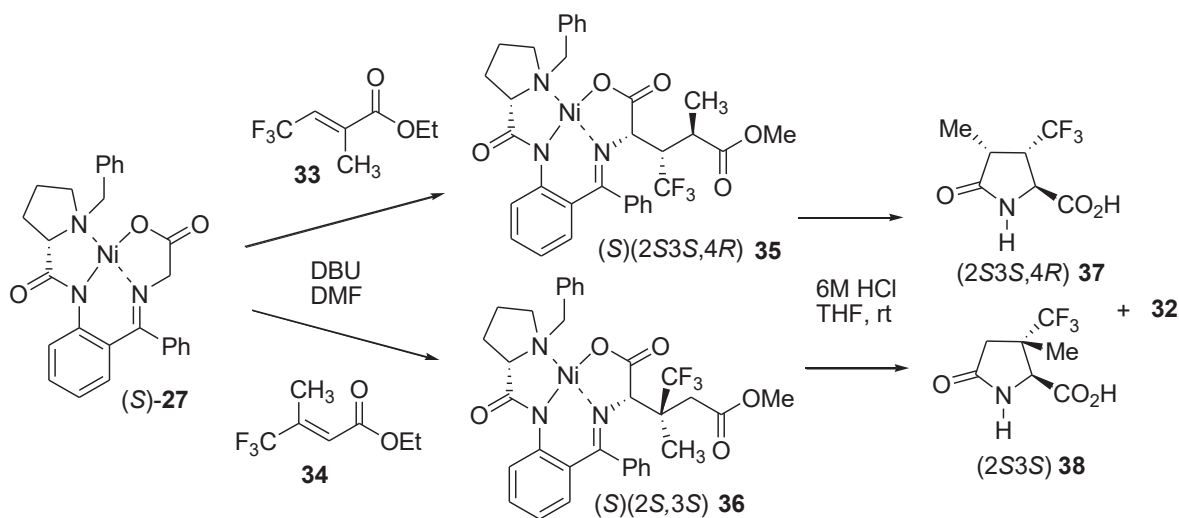
Scheme 5. Michael addition reactions of chiral Ni(II) complex **27**.



Scheme 6. DBU-catalysed Michael addition reactions of chiral Ni(II) complex **27**.

Of particular interest are the reactions between the Ni complex (S)-**27** and bis-substituted Michael acceptors **33** and **34** (Scheme 7) [89]. For instance, the simultaneous formation of α -, β -, and γ -stereogenic centers (eight theoretically possible stereoisomers) in the reaction of complex (S)-**27** with ester **33** represents a challenging stereochemical problem. The Michael addition between (S)-**27** and **33**, conducted in DMF in the presence of DBU, resulted in only two diastereomers (major **35**), in a synthetically useful ratio of 17/1, respectively. The reaction of complex (S)-**27** with

ester **36**, conducted under the same reaction conditions, proceeded with virtually complete diastereoselectivity, furnishing a sole reaction product **36**. Diastereomerically pure products **35** and **36** were disassembled to release interesting bis-substituted pyroglutamic acids **37** and **38**. The high diastereoselectivity in these reactions, particularly the complete stereochemical discrimination between the methyl and trifluoromethyl groups in the addition of (S)-**27**, can be attributed to the steric and electronic effects of the trifluoromethyl group [90, 91].



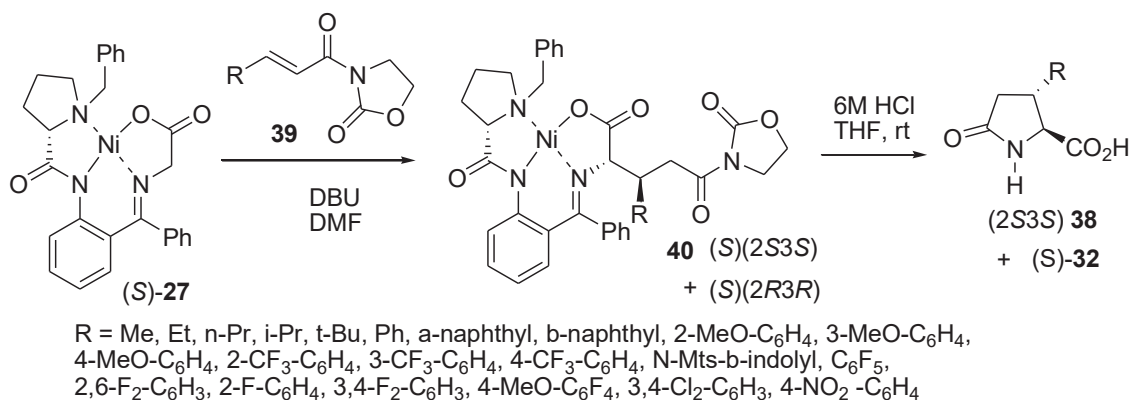
Scheme 7. Michael addition reactions of complex **27** with fluorinated derivatives **33** and **34**.

Interesting results were obtained in the reactions of chiral Ni-complex (S)-**27** [92–94] with oxazolinin-2-one derived Michael acceptors **39** (Scheme 8). These additions were studied using DMF as a solvent and DBU as a base. The reactions were found to occur at high reaction rates with perfect simple diastereoselectivity. On the other hand, the glycine complex enolate *re/si*-face-selectivity was rather poor resulting

generally in mixtures of two diastereomeric products **40**. The highest diastereoselectivity, 5.2/1 ratio of (S)(2S3S) and (S)(2R3R) diastereomers **40**, in the aliphatic series was obtained in the reaction of (S)-**27** with *i*-Pr-containing **39**. Rather interesting and unexpected results were observed in the reactions of (S)-**27** with Michael acceptors bearing aromatic substituents. Thus, the addition between (S)-**27** and

phenyl-containing **39** furnished a mixture of diastereomers **40** in a ratio of 4/1, respectively. In sharp contrast, the reaction between (S)-**27** and pentafluorophenyl-containing **39** occurred almost instantly (<2 min) giving rise to product **40** with excellent diastereoselectivity (ratio > 26/1). Further detailed study of the electronic and steric effects of the substituents on stereochemical outcome in these reactions revealed a general trend that presence of the electron withdrawing substituents on starting **39** led to higher reaction rates and diastereoselectivity, while the

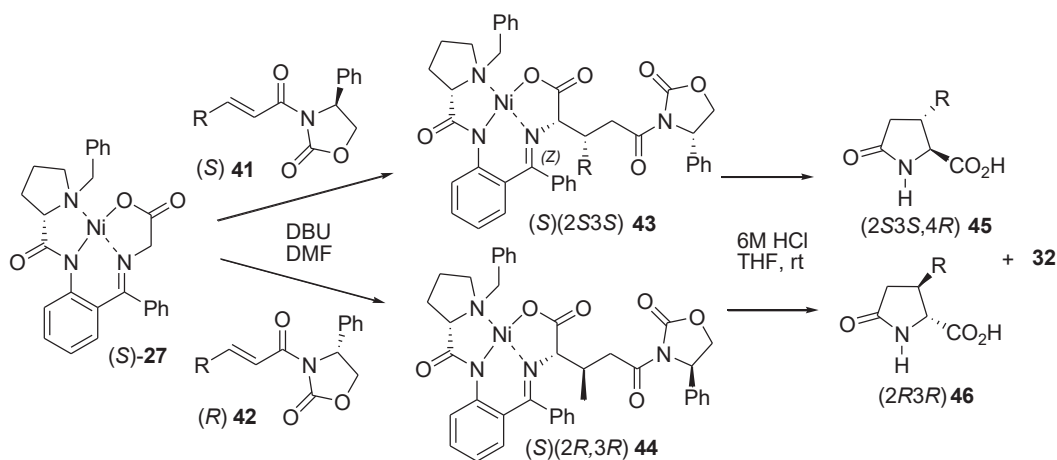
presence of the electron-releasing substituents resulted in low rates and moderate selectivity. To account for these electronic effects dramatically influencing the stereochemical outcome, one can consider electron donor-acceptor attractive interactions between electron deficient aromatic rings, such as pentafluorophenyl [95–97] in particular, and the ketimine phenyl of the starting Ni-complex **27**. The target pyroglutamic acids **38** were obtained after standard disassembly procedure along with the recovery of chiral auxiliary (S)-**32**.



Scheme 8. Michael addition reactions of chiral N(II) complex **27** with oxazolin-2-one derived Michael acceptors **39**.

Considering the strong stereocontrolling effect of oxazolin-2-one chiral auxiliaries [98–100], it was interesting to investigate the corresponding Michael addition reactions using chiral Michael acceptors **41** and **42** (Scheme 9) with the chiral Ni(II) complex **27** [101–103]. The addition reaction between (S)-**27** and crotonyl-derived (S)-**41**, conducted in DMF/DBU at ambient temperature, proceeded to completion at a high rate, furnishing a sole reaction product **43** with the (2S, 3S) configuration of the glutamic acid residue in quantitative chemical yield. This outcome was considered a match case, as both

chiral auxiliaries showed a preference for the α-(S) configuration of the addition product. The reaction of (S)-**27** with crotonyl-derived (R)-**42** was expected to be a mismatch case. The addition was conducted under the same conditions and occurred at a slightly slower rate, giving rise to a sole diastereomerically pure product **44**. The absolute configuration of the glutamic residue in **44** was found to be (2R, 3R). This result was quite unexpected, suggesting that the stereochemical preferences of the chiral Michael acceptor **42** completely overwhelmed those of the chiral Ni-complex (S)-**27**.



Scheme 9. Michael addition reactions between chiral Ni(II) complex **27** and Michael acceptors **41** and **42**.

The same pattern of reactivity and stereochemical preferences was observed in the reactions of (S)-**27** with various Michael acceptors **41** and **42**, bearing, in principle, any alkyl or aromatic substituents (Scheme 9). Of particular interest was the addition between (S)-**27** and the bulky i-Pr-containing **41/42**. When (S)-**41** (R = i-Pr) was used, the reaction proceeded to completion in 30 minutes, affording the product (2S, 3R)-**43** in quantitative chemical yield. In the mismatch case, the addition between (S)-**27** and (R)-**42** (R = i-Pr) was considerably slower and accompanied by the formation of some by-products. Nevertheless, within 4 hours the reaction was completed, and the target product (2R, 3S)-**44** was isolated in 67% yield [101–103]. The results obtained clearly indicated that the steric and electronic nature of the substituents on the starting Michael acceptors **41/42** do not influence the stereochemical outcome of reactions with (S)-**27**, suggesting that the substituents on the C,C double bond of **41/42** are not involved in the stereocontrolling step of the additions.

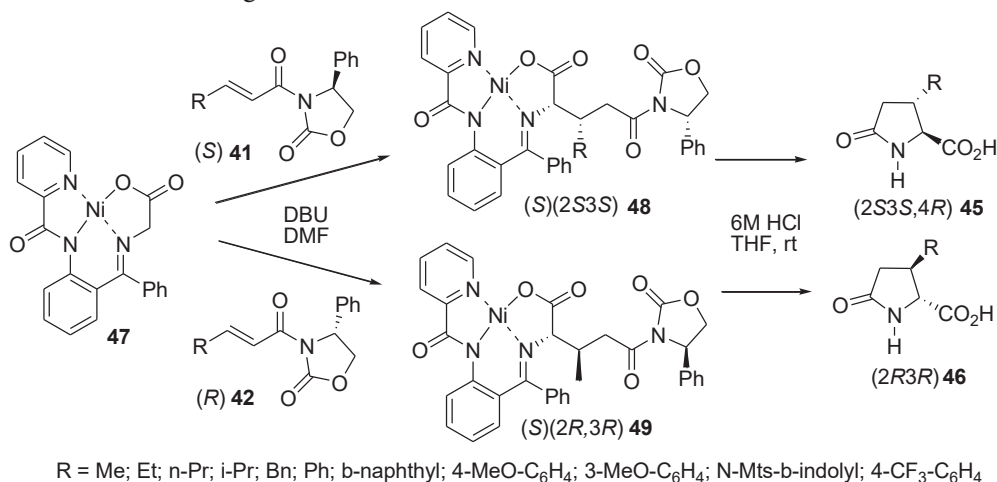
This observation highlighted the generality of the method and underscored the intriguing stereocontrolling power of the chiral oxazolidine-2-one-derived Michael acceptors **41/42** [101–103]. The final amino acids **45** and **46** were prepared by acidic disassembly of the addition products, along with the recovery and reuse of the chiral ligand **32**.

To take advantage of this discovery, the reactions of achiral Ni(II) complex **47** (Scheme 10) with chiral Michael acceptors **41/42** were investigated [104–106]. Picolinic acid-derived achiral complex **47** [107] was reacted with Michael acceptor **42** in DMF at room temperature in the presence of 15 mole % DBU. The addition occurred at a high rate, affording a sole reaction product **48** in quantitative chemical yield. Application of (S)-configured **41** mirrored the result obtained for (R)-**42**, affording individual product **48** containing the (2S, 3S)-enantiomer of β -methyl glutamic acid. The same excellent stereochemical outcome was observed in the reaction of all alkyl-containing **41/42** (R = Et, n-Pr, Bn), except for the addition between

complex **47** and bulky *i*-Pr-containing **41/42**. In this case, the reaction proceeded very slowly, allowing for about 30% conversion of the starting compounds in 4 hours. Though the corresponding products **48** and **49** were isolated in 15% yield, these results suggested that the present method could not be extended to substrates containing tertiary alkyl groups.

To examine the applicability of the method to an aromatic series, which would lead to the synthesis of the corresponding 3-aryl substituted amino acids, the reactions of complex **47** with (*S*)- and (*R*)-configured Michael ac-

ceptors **41/42** containing classical phenyl and naphthyl groups, as well as derivatives bearing a phenyl ring with electron-withdrawing and electron-donating substituents, were studied. In all cases, regardless of the steric or electronic nature of the substituent on the starting **41/42**, major products **48** or **49** were obtained with at least 95% de and excellent chemical yields. In both aliphatic and aromatic series, the stereochemical outcome was shown to be kinetically controlled, affording products with relative topicity like.



Scheme 10. Michael additions between achiral Ni(II) complex **47** and chiral acceptors **41** and **42**.

To demonstrate the synthetic efficiency of this method, >25 g of enantiomerically pure β -phenyl pyroglutamic acid (2*S*, 3*R*)-**45** was prepared in >85% total yield in less than 14 hours. In this case, the crystalline product **49** of the addition between **47** and (*R*)-**42** was isolated simply by filtration after pouring the reaction mixture into water. Without additional purification, product **49** was disassembled to afford the target amino acid **46**.

The relatively broad substrate generality, excellent chemical yields and diastereoselectivity,

combined with the simplicity of the experimental procedures, render the present method synthetically superior to the literature method for preparing various 3-substituted pyroglutamic acids and related amino acids.

Apparent success of these Ni(II) complexes in the asymmetric Michael addition reactions stimulated interest in the developing of new types of chiral as well as achiral Ni(II) complexes [108–111]. Figure 4 presents just a few most prospective types.

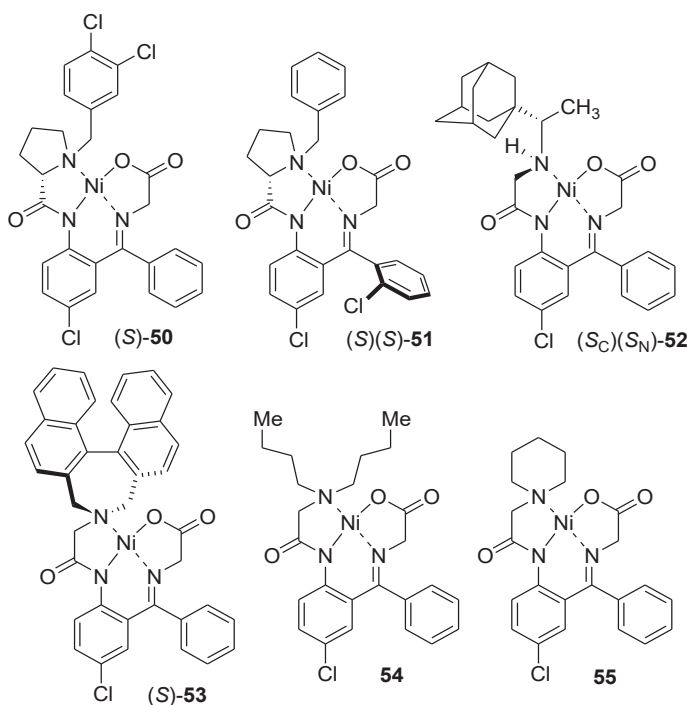


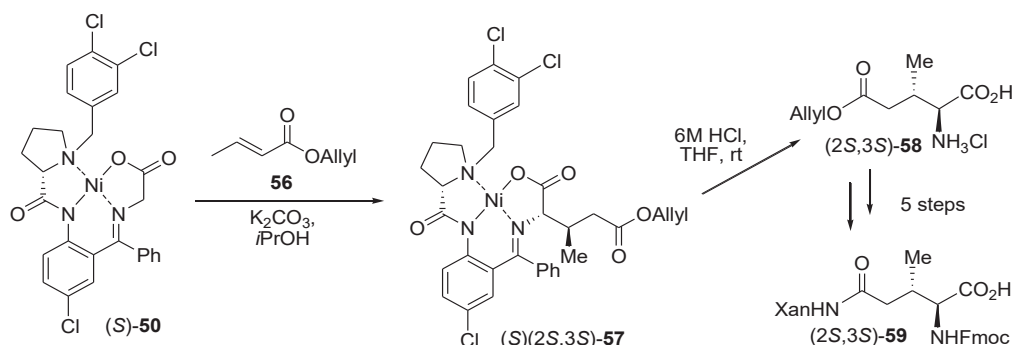
Fig. 4. New generation of chiral and achiral Ni(II) complexes.

For example, chlorinated Ni(II) complex **50**, available in both enantiomeric forms [112] possesses exceptional stereocontrolling properties [113,114] due to electrostatic interactions between electron-rich benzophenone phenyl and electron-deficient chlorine-containing rings [115]. Thus, complex (S)-**50** was successfully used in Michael addition for the synthesis of (2S, 3S)-3-methylglutamine, the key compound used in the total synthesis of cytotoxic marine peptides callipeltin O and Q [116]. As presented in Scheme 11, glycine Ni(II) complex **50** was reacted with Michael acceptor **51** under specially designed conditions to keep intact the allyl ester moiety [117].

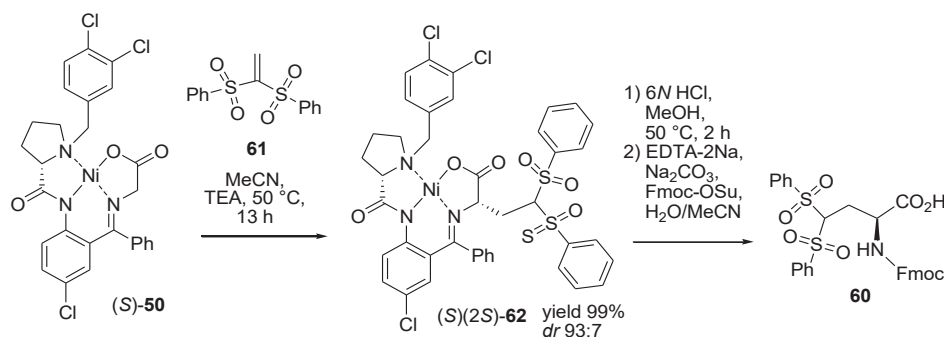
The acidic disassembly of the major diastereomeric product (S)(2S3S)-**57** was also performed under strictly controlled conditions using THF as a solvent. Allyl ester hydrochloride **58** was subsequently transformed into the target properly protected for peptide

synthesis glutamine derivative **59** in five steps.

It was demonstrated that the substitution of phenylalanine in endogenous peptides with a bulkier and more lipophilic β -phenylphenylalanine (diphenylalanine, DPA) usually leads to improved binding to the apolar site of the targeted receptors. In particular, DPA-modified peptides were found to possess enhanced biological profiles in the series of thrombin inhibitors [118], angiotensin-converting enzyme (ACE) inhibitors [119], HIV protease inhibitors [120], pain-related norepinephrine transporter inhibitors [121], μ and δ opioid receptors [122]. In this regard, 2-amino-4,4-bis-(phenylsulfonyl)butanoic acid **60** (Scheme 12) is one of the promising structures featuring enhanced steric bulk and lipophilicity. Chiral Schiff base complex **50** was selected for preparation of this amino acids via Michael addition using specially designed vinyl-disulfonyl Michael acceptor **61** [123, 124].



Scheme 11. Synthesis of properly protected (2S,3S)-3-methylglutamine **59** using rationally designed chlorinated Ni(II)-complex (S)-**50**.

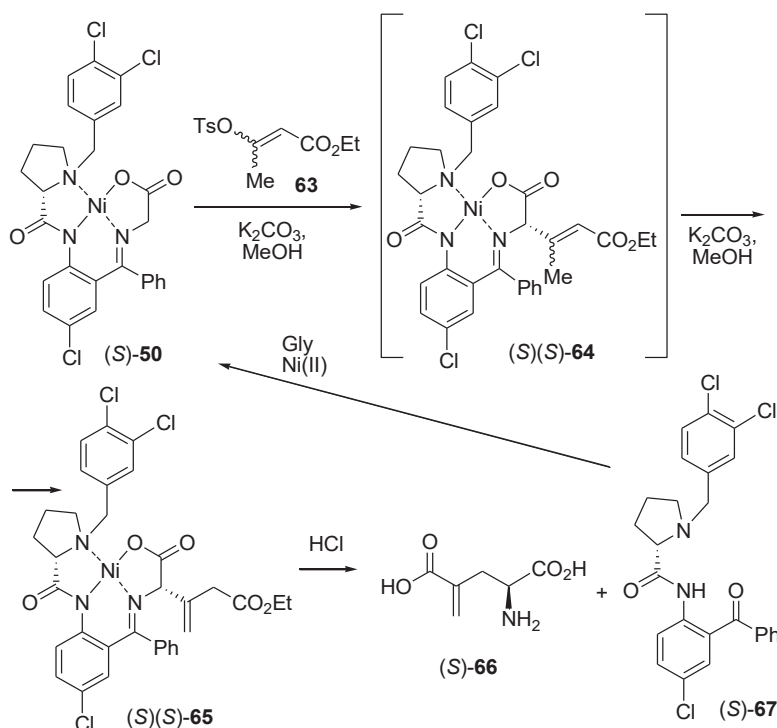


Scheme 12. Asymmetric synthesis of sterically bulky and lipophilic 2-amino-4,4-bis-(phenylsulfonyl)butanoic acid (S)-**60**.

It was found that due to the high electrophilicity of reagent **61**, the corresponding Michael addition with chiral glycine complex **50** can be efficiently conducted using triethylamine as base. Optimization of the reaction conditions allowed for preparation of the diastereomeric products with an excellent yield (99%) and excellent diastereoselectivity (93/7). The major product **62** was transformed into the corresponding Fmoc derivative of 2-amino-4,4-bis-(phenylsulfonyl)butanoic acid **60** under standard conditions.

Rather unexpected results were reported for the Michael addition between chiral glycine Ni(II) complex **50** and Michael acceptor **63** bearing a tosylate leaving group (Scheme 13)

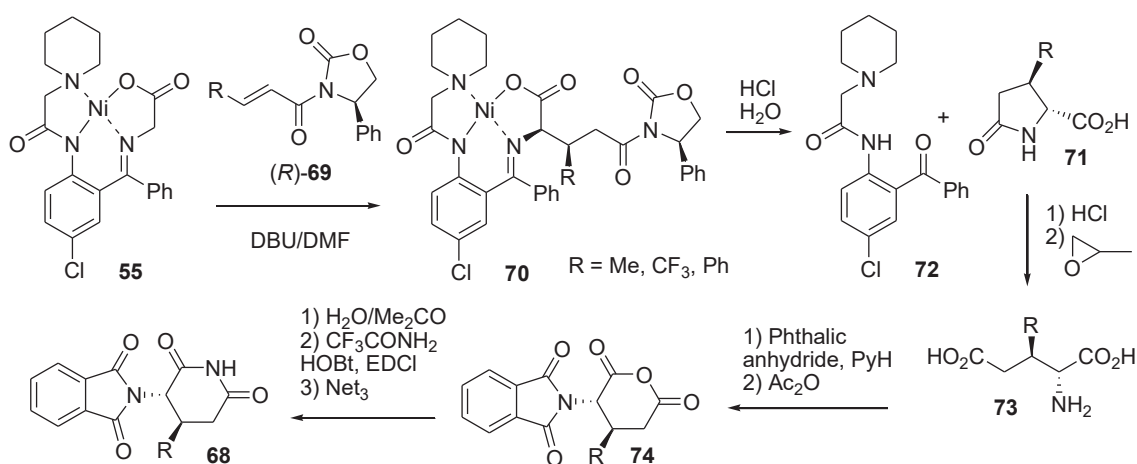
[125]. The original goal of this research was preparation of addition product **64** containing the corresponding β -methyl-unsaturated amino acid moiety. However, quite surprisingly, compound **65** was isolated as the major reaction product. (S)-3-Methyleneglutamic acid **66** was previously unknown in enantiomerically pure form. One can assume that the target addition product **64** underwent base-catalyzed isomerization to **65**. Major product (S) (S)-**65** was isolated in diastereomerically pure form and disassembled to release the unsaturated amino acid (S)-**66** along with the chiral ligand (S)-**67**, which was recycled and reused for preparation of new portions of starting chiral glycine Schiff base Ni(II) complex (S)-**50**.



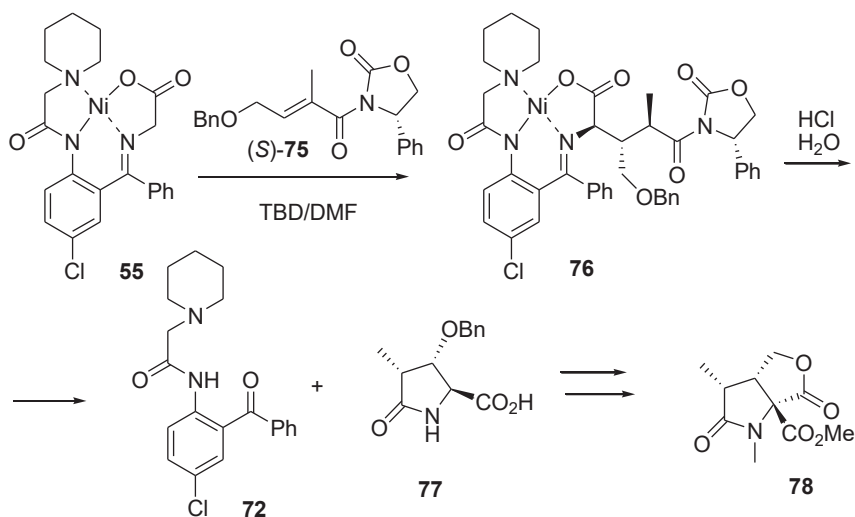
Scheme 13. Asymmetric synthesis of (S)-3-methyleneglutamic acid **66** via Michael addition reaction.

Achiral complex **55** was selected for preparation of 4-substituted configurationally stable thalidomide derivatives **68** (Scheme 14) [126,127]. As illustrated in Scheme 14, key step in the multistep synthesis of the thalidomide derivatives is Michael addition reaction of achiral glycine Schiff base derivatives **55** with chiral (*R*)- or (*S*)-*N*-(*E*-enoyl)-4-phenyl-1,3-oxazolidine-2-one **69** were conducted in DMF using DBU as a base. The resultant addition products **70** were isolated as individual diastereoisomers in quantitative chemical yields. Products **70** can be easily hydrolyzed under mild conditions and, upon a workup procedure, transformed to the pyroglutamic acids **71** along with recovery of ligand **72** which can be used for preparation of the starting complex **55**. Hydrolysis of pyroglutamic acids **71** with 3N HCl at reflux afforded 3-substituted glutamic acids **73** in high chemical yield.

The next step, transformation of acid **73** to *N*-phthaloyl anhydrides **74**, was an important step as it was expected that during this process the corresponding alpha-stereogenic carbon would undergo epimerization setting up the trans-configuration in the final products **74** [128]. Indeed the transformation of acids **73** to *N*-phthaloyl anhydrides **74**, occurred as expected furnishing a single diastereomers **74** in up to 70% (two steps) yield. The final transformation of enantio- and diastereomerically pure **74** to the target 4-substituted thalidomides **68**, was conducted in two major steps under the standard conditions [129]. After hydrolysis of anhydrate **74** with water in acetone, the corresponding intermediates were smoothly transformed to **68** using trifluoroacetamide as a source of nitrogen. The target products **68** were isolated in good yields (~70% for two steps) as single diastereomers.



Scheme 14. Asymmetric synthesis of 4-substituted thalidomide derivatives via Michael addition reactions.



Scheme 15. Asymmetric synthesis of 3,4-disubstituted pyroglutamic acid **77** via Michael addition reaction of achiral Ni(II) complex **55**.

With the goal of developing a new approach for preparing analogs of oxazolomycin family products **78** (Scheme 15), the reaction of achiral Ni(II) complex **55** with disubstituted chiral Michael acceptor **75** was considered a key step, allowing the formation of the structural frame as well as the three required stereogenic centers. Interestingly, the reaction of

55 with **75** conducted in DMF/DBU did not proceed at all, resulting in complete recovery of the starting material. Eventually, after numerous attempts to find the right conditions, 1,5,7-triazabicyclo [4.4.0] dec-5-ene (TBD), slightly more basic than DBU, was found to catalyze the reaction, affording a mixture of the Michael addition products. It was determined

that the reaction mixture consisted of six diastereomers obtained in a ratio of 68:8:7:7:7:3. The major product was separated by silica gel column chromatography to give **76** in 62% isolated yield [130].

Under standard acidic conditions, the major diastereomer **76** was disassembled to release the target pyroglutamic acid **77** along with the reusable tridentate ligand **72**. Key amino acid **77** was then transformed into oxazolomycin analog **78** using literature procedures.

Application of this Ni(II) complex methodology is not limited to glutamic acids and can be extended to preparing various other derivatives using, to mention just a few, alkyl halide alkylations [131–133], aldol [134–136], Mannich [113, 137] addition reactions and DKR [138, 139].

CONCLUSIONS. This review has sought to emphasize the key synthetic and methodological concepts developed to control both simple and facial stereoselectivity in Michael addition reactions involving nucleophilic glycine equivalents and α , β -unsaturated carboxylic acid derivatives. While some methods discussed hold historical significance, others present practical, generalized, and synthetically efficient solutions. However, various synthetic and stereochemical challenges remain, particularly in reactions involving α -, α , β -bis-, and β , β -bis-substituted α , β -unsaturated carboxylic acid derivatives. As the importance of sterically constrained glutamic/pyroglutamic acids and related amino acids continues to grow, Michael addition reactions—offering the most straightforward and generalized approach to this family of amino acids—will remain a fascinating and challenging area in the field of asymmetric synthesis.



We gratefully acknowledge the financial support from the National Natural Science Foundation of China (No. 21761132021) and the Qing-Lan Project of Jiangsu Province (for Han) and IKERBASQUE, Basque Foundation for Science (for Soloshonok).

АСИМЕТРИЧНИЙ СИНТЕЗ ХІРО-ОБМЕЖЕНИХ ГЛУТАМІНОВИХ КИСЛОТ І СПОРІДНЕНИХ СПЛУК ЗА ДОПОМОГОЮ РЕАКЦІЙ МІХАЕЛЯ

*Аліція Взорек, Олександр Е. Сорочинський,
Карел Д. Кліка, Тайзо Оно, Цзяньлін Хань,
Вадим А. Солошонов*

¹Інститут хімії, Університет імені Яна Кохановського в Кельце,

вул. Університетська 7, 25–406 Кельце, Польща;

²Відділ тонкого органічного синтезу, Інститут біоорганічної хімії та нафтохімії ім. В. П. Кухаря, Національна академія наук України,

вул. Мурманська 1, Київ 02094, Україна;

³Аналіз молекулярних структур, Німецький центр дослідження раку (DKFZ), Ім-Нойенгеймер-Фельд 280, 69120 Гайдельберг, Німеччина;

⁴Національний інститут передових промислових наук і технологій, 463–8560, Нагоя, Японія;

⁵Цзянсуський центр спільних інновацій з ефективного оброблення та використання лісових ресурсів, Коледж хімічної інженерії, Нанкінський лісотехнічний університет, Нанкін 210037, Китай;

⁶Кафедра органічної хімії I, Хімічний факультет, Університет Країни Басків UPV/EHU, проспект Мануеля Лардисабала 3, 20018 Сан-Себастьян, Іспанія;

⁷IKERBASQUE, Баскський фонд науки, вул. Марії Діасде Харо 3, площа Бізкайя, 48013 Більбао, Іспанія.

email: vadimsoloshonok@gmail.com

Реакції Міхаеля, що включають нуклеофільні еквіваленти гліцину та α , β -ненасичені похідні карбонових кислот, пропонують короткий та узагальнений методологічний підхід до синтезу сімейства амінокислот з α -обмеженням, що містять п'ять атомів вуглецю. Ці амінокислоти відіграють вирішальну роль у дизайні нових пептидів та в з'ясуванні тривимірної структури пептидів/білків та їхніх біологічних функцій/активності. У цьому огляді узагальнено значні синтетичні та методологічні досягнення у зазначеній галузі на сьогодні. Кожен розглянутий метод включає оцінку синтетичних можливостей і обмежень, практичності та ефективності процедур і механістичне обґрунтування спостережуваних стереохімічних переваг.

Ключові слова: асиметричний синтез, реакції Міхаеля, еквіваленти гліцину, металлокомплекси, шіфові основи, конформації, стеричне обмеження, глутамінова кислота, піроглутамінова кислота, пролін.

REFERENCES

1. Vickery H. B., Schmidt C. L. A. The history of the discovery of the amino acids. *Chem. Rev.* 1931. **9**: 169–318.
2. Fieulaine S., Boularot A., Artaud I., *et al.* Trapping Conformational States Along Ligand-Binding Dynamics of Peptide Deformylase: The Impact of Induced Fit on Enzyme Catalysis. *PLOS Biology*. 2011. **9**: e1001066.
3. Kaiser A., Coin I. Capturing Peptide–GPCR Interactions and Their Dynamics. *Molecules*. 2020. **25**: 4724.
4. Rosenblum D., Joshi N., Tao W. *et al.* Progress and challenges towards targeted delivery of cancer therapeutics. *Nature Communications*. 2018. **9**: 1410.
5. Mitchell M. J., Billingsley M. M., Haley R. M. *et al.* Engineering precision nanoparticles for drug delivery. *Nature Reviews Drug Discovery*. 2021. **20**: 101–124.
6. Muttenthaler M., King G. F., Adams D. J., Alewood, P. F. Trends in peptide drug discovery. *Nature Reviews Drug Discovery*. 2021. **20**: 309–325.
7. Han J., Konno H., Sato T. *et al.* Tailor-made amino acids in the design of small-molecule blockbuster drugs. *Eur. J. Med. Chem.* 2021. **220**: 113448. doi.org/10.1016/j.ejmech.2021.113448.
8. Liu J., Han J., Izawa K. *et al.* Cyclic tailor-made amino acids in the design of modern pharmaceuticals. *Eur. J. Med. Chem.* 2020. **208**: 112736. Doi:10.1016/j.ejmech.2020.112736.
9. Mei H., Han J., White S. *et al.* Tailor-made amino acids and fluorinated motifs as prominent traits in modern pharmaceuticals. *Chem.–Eur. J.* 2020. **26**: 11349–11390. https://doi.org/10.1002/chem.202000617.
10. Ma J.S. Unnatural amino acids in drug discovery. *Chim Oggi–Chem Today*. 2003 **21**: 65–68.
11. Fosgerau K., Hoffmann T. Peptide therapeutics: Current status and future directions. *Drug Discov Today*. 2015. **20**: 122–128.
12. Asymmetric Synthesis and Application of α -Amino Acids. ACS Symposium Series #1009, Soloshonok V.A., Izawa K. Eds. Oxford University Press. 2009.
13. Hodgson D.R.W., Sanderson J.M. The synthesis of peptides and proteins containing non-natural amino acids. *Chem. Soc. Rev.* 2004. **33**: 422–430.
14. Henninot A., Collins J. C., Nuss J. M. The current state of peptide drug discovery: Back to the future? *J. Med. Chem.* 2018. **61**: 1382–1414.
15. Mei H., Han J., Klika K. D. *et al.* Applications of fluorine containing amino acids for drug design. *Eur. J. Med. Chem.* 2020. **186**: 111826. https://doi.org/10.1016/j.ejmech.2019.111826.
16. Craik D. J., Fairlie D. P., Liras S., Price D. The future of peptide based drugs. *Chem. Biol. Drug. Des.* 2013. **81**: 136–147.
17. Han J., Sorochinsky A. E., Ono T. *et al.* Biomimetic Transamination – a Metal-Free Alternative to the Reductive Amination. Application

- for Generalized Preparation of Fluorine-Containing Amines and Amino Acids. *Cur. Org. Synthesis* 2011. **8**: 281–294.
18. Stevenazzi A., Marchini M., Sandrone G. *et al.* Amino acidic scaffolds bearing unnatural side chains: An old idea generates new and versatile tools for the life sciences. *Bioorg. Med. Chem. Lett.* 2014. **24**: 5349–5356.
19. Han J., Konno H., Sato T. *et al.* Peptidomimetics and Peptide-Based Blockbuster Drugs. *Curr. Org. Chem.* 2021. **25**: 1627–1658. DOI: 10.2174/1385272825666210610155047
20. Han J., Wzorek A., Dhawan G. *et al.* New drugs appearing on the market in 2023: molecules containing fluorine and fragments of tailor-made amino acids. *Ukr. Bioorg. Acta* 2024. **19** (1): 3–20. <https://doi.org/10.15407/bioorganica2024.01.003>
21. Pagire S. H., Lee E., Pagire H. S. *et al.* Design, synthesis and biological evaluation of glutamic acid derivatives as anti-oxidant and anti-inflammatory agents. *Bioorg. Med. Chem. Lett.* 2017. **28**: 5290–5321.
22. Panday S. K. Pyroglutamic Acid and its Derivatives: The Privileged Precursors for the Asymmetric Synthesis of Bioactive Natural Products. Bentham Science. 2020. **17**(6): 626–6462.
23. Sun M., Han L., Li A. *et al.* Design, synthesis, and evaluation of novel L-pyroglutamic acid derivatives as potent antifungal agents. *Chemistry of Natural Compounds*. 2021. **57**(6): 10814.
24. Meldrum B. S. Glutamate as a neurotransmitter in the brain: Review of physiology and pathology. *Journal of Nutrition*. 2000. **130**: 1007S–1015S.
25. Luo W., Fang X., Wang C. *et al.* Terminus-immobilization effect on peptide conformations and peptide-peptide interactions. *Nano Research*. 2023. **16**: 13498–13508.
26. Hruby V. J. Conformational restrictions of biologically active peptides via amino acid side chain groups *Life Sci.* 1982. **31**: 189–199.
27. Balaram P., Ramaseshan S., Eds. Molecular Conformation and Biological Interactions. Indian Academy of Science, Bangalore. 1991.
28. Ramachandran G.N., Sasisekharan V. Conformation of polypeptides and proteins. *Adv Protein Chem.* 1968. **23**: 283–438. doi: 10.1016/s0065-3233(08)60402-7.
29. Scheraga H.A. Theoretical and experimental studies of conformations of polypeptides. *Chem. Rev.* 1971. **71**: 195–217. doi: 10.1021/cr60270a003.
30. Bloom S. M., Fasman G. D., deLoze C., Blout E. R. The Effect of Amino Acid Composition on the Conformations of Synthetic Polypeptides, Polymers and Copolymers of L-Methionine S-Methyl-L-cysteine and L-Valine. *J. Am. Chem. Soc.* 1962. **84**: 458–463. DOI: 10.1021/ja00862a027.
31. Gibson S. E., Guillo N., Tozer M. J. Towards Control of χ -Space: Conformationally Constrained Analogues of Phe, Tyr, Trp, and His. *Tetrahedron*. 1999. **55**: 585–601. DOI: 10.1002/CHIN.199923302
32. Ellis T. K., Martin C. H., Tsai G. M. *et al.* Efficient Synthesis of Sterically Constrained Symmetrically α,α -Disubstituted α -Amino Acids under Operationally Convenient Conditions. *J. Org. Chem.* 2003. **68**: 6208–6214.
33. Soloshonok V. A., Sorochinsky A. E. Practical Methods for the Synthesis of Symmetrically α,α -Disubstituted- α -Amino Acids, *Synthesis* 2010. **14**: 2319–2344.
34. Ellis T. K., Martin C. H., Ueki H., Soloshonok V. A. Efficient, Practical Synthesis of Symmetrically α,α -Disubstituted α -Amino Acids. *Tetrahedron Lett.* 2003. **44**: 1063–1066.
35. Sato T., Izawa K., Aceña J. L. *et al.* Tailor-Made α -Amino Acids in Pharmaceutical Industry: Synthetic Approaches to (1R,2S)-1-Amino-2-vinylcyclopropane-1-carboxylic Acid (Vinyl-ACCA). *Eur. J. Org. Chem.* 2016. 2757–2774. DOI: 10.1002/ejoc.201600112.
36. Taylor S. M., Yamada T., Ueki H., Soloshonok V. A. Asymmetric Synthesis of Enantiomerically Pure 4-Aminoglutamic Acids via Methylenedimerization of Chiral Glycine Equivalents with Dichloromethane under Operationally Convenient Conditions. *Tetrahedron Lett.* 2004. **45**: 9159–9162.

37. Ellis T. K., Hochla V. M., Soloshonok V. A. Efficient Synthesis of 2-Aminoindane-2-Carboxylic Acid via Dialkylation of Nucleophilic Glycine Equivalent. *J. Org. Chem.* 2003. **68**: 4973–4976.
38. Hruby V. J., Li G., Haskell-Luevano C., Shenderovich M. D. Design of peptides, proteins, and peptidomimetics in chi space. *Biopolymers*. 1997. **43**: 219–266.
doi: 10.1002/bip.10970282.
39. Qiu W., Gu X., Soloshonok V. A. *et al.* Stereoselective synthesis of conformationally constrained reverse turn dipeptide mimetics. *Tetrahedron Lett.* 2001. **42**: 145–148.
40. Hruby V.J. Conformational and topographical considerations in the design of biologically active peptides. *Biopolymers*. 1993. **33**: 1073–1082.
doi: 10.1002/bip.360330709.
41. Cai M., Cai C., Mayorov A. V. *et al.* Biological and conformational study of β -substituted prolines in MT-II template: steric effects leading to human MC5 receptor selectivity. *J. Pept. Res.* 2004. **63**: 116–131.
42. Hruby V. J., Al-Obeidi F., Kazmierski W. M. Emerging approaches in the molecular design of receptor-selective peptide ligands: conformational, topographical and dynamic considerations. *Biochemical J.* 1990. **268**: 249–262.
doi: 10.1042/bj2680249.
43. O'Donnell M. J., Bennett W. D. Wu S. The stereoselective synthesis of α -amino acids using phase-transfer catalysis. *Tetrahedron*. 2013. **69**(2): 123–130.
44. Kim Y., Park J., Kim M.J. Dynamic kinetic resolution of amines and amino acids by enzyme-metal cocatalysis. *Chem. Cat. Chem.* 2011. **3**: 271–277.
45. Takeda R., Kawamura A., Kawashima A. *et al.* Chemical Dynamic Kinetic Resolution and (S)/(R)-Interconversion of Unprotected α -Amino Acids. *Angew. Chem., Int. Ed.* 2014.
DOI: 10.1002/anie.201407944.
46. So S.M., Kim H., Mui L., Chin J. Mimicking nature to make unnatural amino acids and chiral diamines. *Eur. J. Org. Chem.* 2012. 229–241.
47. D'Arrigo P., Servi S. *et al.* Synergy between catalysts: enzymes and bases. *Catal. Sci. Technol.* 2012. **2**: 1606–1616.
48. Bera K., Namboothiri I. Asymmetric synthesis of quaternary α -amino acids and their phosphonate analogues. *Asian J. Organ. Chem.* 2014. **3**: 1234–1260.
49. He G., Wang B., Nack W.A., Chen, G. Syntheses and transformations of α -amino acids via palladium-catalyzed auxiliary-directed sp^3 C-H functionalization. *Acc. Chem. Res.* 2016. **49**: 635–645.
50. Wang Y., Song X., Wang J. *et al.* Recent approaches for asymmetric synthesis of α -amino acids via homologation of Ni(II) complexes. *Amino Acids*. 2017. **49**: 1487–1520.
DOI: 10.1007/s00726-017-2458-6.
51. Sorochinsky A. E., Aceña J. L., Moriwaki H. *et al.* Asymmetric synthesis of α -amino acids via homologation of Ni(II) complexes of glycine Schiff bases; Part 1: Alkyl halide alkylations. *Amino Acids*. 2013. **45**: 691–718.
DOI: 10.1007/s00726-013-1539-4.
52. Sorochinsky A. E., Aceña J. L., Moriwaki H. *et al.* Asymmetric synthesis of α -amino acids via homologation of Ni(II) complexes of glycine Schiff bases. Part 2: Aldol, Mannich addition reactions, deracemization and (S) to (R) interconversion of α -amino acids. *Amino Acids*. 2013. **45**: 1017–1033.
DOI 10.1007/s00726-013-1580-3.
53. O'Donnell M. J., Boniece J. M. Enantioselective synthesis of α -amino acids via phase-transfer catalysis. *Tetrahedron*. 2015. **71**(4): 456–462.
54. Nian Y., Wang J., Zhou S. *et al.* Recyclable Ligands for the Non-Enzymatic Dynamic Kinetic Resolution of Challenging α -Amino Acids. *Angew. Chem., Int. Ed.* 2015. **54**: 12918–12922.
DOI: 10.1002/anie.201507273.
55. O'Donnell M. J., Earp S. E. Advances in the asymmetric synthesis of α -amino acids. *Tetrahedron* 2017. **73**(5): 567–575.
56. Soloshonok V. A., Kirilenko A. G., Fokina N. A. *et al.* Chemo-Enzymatic Approach to the Synthesis of Each of the Four Isomers of α -Alkyl- β -Fluoroalkyl-Substituted β -Amino Acids. *Tetrahedron: Asymmetry*. 1994. **5**: 1225–1228.

57. O'Donnell M. J., Wu S. Phase-transfer catalysis in the synthesis of α -amino acids: Recent developments. *Tetrahedron*. 2019. **75**(3): 432–440.
58. O'Donnell M. J., Bennett W. D. Stereocontrolled synthesis of α -amino acids using phase-transfer catalysis. *Tetrahedron*. 2021. **77**(2): 289–297.
59. Soloshonok V. A., Ono T., Soloshonok I. V. Enantioselective Biomimetic Transamination of β -Keto Carboxylic Acid Derivatives. An Efficient Asymmetric Synthesis of β -Fluoroalkyl- β -Amino Acids. *J. Org. Chem.* 1997. **62**: 7538–7539.
60. Shioiri T., Hamada, U. Efficient Syntheses of Biologically Active Peptides of Aquatic Origin Involving Unusual Amino Acids, *Synlett*. 2001. 184–201.
DOI: 10.1055/s-2001-10758.
61. Han J., Sorochinsky A. E., Ono T. *et al.* Biomimetic Transamination – a Metal-Free Alternative to the Reductive Amination. Application for Generalized Preparation of Fluorine-Containing Amines and Amino Acids. *Cur. Org. Synthesis*. 2011. **8**: 281–294.
62. Cativiela C., Diaz-de-Villegas M. D. Stereoselective Synthesis of Quaternary Alpha-Amino Acids. Part 2: Cyclic Compounds. *Tetrahedron: Asymmetry*. 2000. **11**: 645–698.
DOI 10.1016/j.tetasy.2000.11.001.
63. Abellan T., Chinchilla R., Galindo N. *et al.* New Oxazinone and Pyrazinone Derivatives as Chiral Reagents for the Asymmetric Synthesis of α -Amino Acids. *Eur. J. Org. Chem.* 2000. 2689–2698.
DOI:10.1002/CHIN.200039248.
64. Shibata N., Nishimine T., Shibata N., *et al.* Organic base-catalyzed stereodivergent synthesis of (R)- and (S)-3-amino-4,4,4-trifluorobutanoic acids. *Chem. Commun.* 2012. **48**: 4124–4126.
DOI: 10.1039/C2CC30627A.
65. Schöllkopf U., Grioth U., Deng C. Enantioselective Synthesis of (R)- α -Vinylamino Acids. *Angew. Chem. Int. Ed. Engl.* 1981. **20**: 798–803.
<https://doi.org/10.1002/anie.198109771>.
66. Schöllkopf U., Pettig D., Busse U. Asymmetric Synthesis via Heterocyclic Intermediates; XXXI: Asymmetric Synthesis of Glutamic Acids and Derivatives thereof by the Bislactim-Ether Method. *Synthesis* 1986. 737–740.
DOI 10.1055/s-1986-31760.
67. Hartzoulakis B., Gani D. J. Syntheses of (2S,3R)- and (2S,3S)-3-methylglutamic Acid. *Chem. Soc. Perkin Trans.* 1994. 2525–2529.
DOI 10.1039/CT994122525.
68. Cahn R. S., Ingold C. K., Prelog V. Specification of Molecular Chirality. *Angew. Chem., Int. Ed.* 1966. **5**: 385–415.
69. Hartwig W., Born L. Diastereoselective and enantioselective total synthesis of the hepatoprotective agent clausenamides. *J. Org. Chem.* 1987. **52**: 4352–4358.
70. Pettig D., Schöllkopf U. Asymmetric Synthesis via Heterocyclic Intermediates; XXXVIII: Asymmetric Synthesis of Dimethyl (R)-2-Amino-(E)-hept-4-enedioates by the Bislactim Ether Method. *Synthesis*. 1988. 173–175.
71. Suzuki K., Seebach D. threo-3-Alkyl-and-Arylglutamic Acid Derivatives by Michael Additions of Boc-BMI Li-Enolates to 2, 6-Di-*t*-butyl-4-methoxyphenyl Alkenoates on the Diastereoselectivity of the Coupling of Trigonal Centers Involving Heterocyclic Li-Enolates. *Liebigs Annalen der Chemie*. 1992. **13**(1): 51–61.
72. Schlecker R., Seebach D., Lubosch W. CH-acidity in the α -position to the N-atom in N,N-dialkylamides with a sterically protected carbonyl group for nucleophilic aminoalkylation. *Helv. Chim. Acta*. 1978. **61**(1): 512–526.
73. Seebach D., Locher R. α , β -Unsaturated Carbonyl Compounds with Sterically Protected Carbonyl Groups–Enforced a_3 versus a_1 Reactivity. *Angew. Chem. Int. Ed.* 1979. **18**(12): 957–958.
74. Seebach D., Häner R., Vettiger T. Nucleophilic ring opening of α -nitrocyclopropanecarboxylic acid-arylesterns with sterically protected but electronically active carbonyl and nitro groups. A new principle of α -amino acid synthesis (2-aminobutanoic acid a_4 -synthon). *Helv. Chim. Acta*. 1987. **70**(6): 1507–1515.
75. Seebach D., Vettiger T., Müller H. M. *et al.* Stereoselective hydroxyalkylations of (S)-2-azeti-

- dinecarboxylic acid. *Liebigs Annalen der Chemie*. 1990. 7: 687–95.
76. Seebach D., Ertas M., Locher R., Bernd Schweizer W. Trityl ketone and trityl enone. Contributions to sterically enforced Michael addition and diastereoselective aldol addition. *Helv. Chim. Acta*. 1985. **68** (1): 264–82.
77. Cooke Jr MP. Sterically directed conjugate addition reactions of unsaturated esters. *J. Org. Chem.* 1986. **51**(9): 1637–1638.
78. Kanemasa S., Tatsukawa A., Wada E. Highly diastereoselective Michael addition of lithiated camphor imines of glycine esters to. Alpha-, beta-unsaturated esters. Synthesis of optically pure 5-oxo-2, 4-pyrrolidinedicarboxylates of unnatural stereochemistry. *J. Org. Chem.* 1991. **56**(8): 2875–83.
79. Soloshonok V. A., Kacharov A. D., Avilov D. V., *et al.* Transition Metal/Base-Catalyzed Aldol Reactions of Methyl α -Isocyanoacetate with Prochiral Ketones, a Straightforward Approach to Stereochemically Defined β , β -Disubstituted- β -Hydroxy- α -Amino Acids. Scope and Limitations. *J. Org. Chem.* 1997. **62**: 3470–3479.
80. Belokon Y. N., Bulychev A. G., Ryzhov M. G. *et al.* General method of diastereo- and enantioselective synthesis of β -hydroxy- α -amino acids by condensation of aldehydes and ketones with glycine. *J. Chem. Soc. Perkin Trans. I*. 1986. 1865–1877.
DOI:10.1021/ja00300a030.
81. Belokon Y. (s)-2-[N-(N'-Benzylpropyl) Amino] Benzophenone (BPB)-A Reagent for the Synthesis of Optically Pure α -Amino Acids. *Janssen Chim. Acta*. 1992. **10**: 4–15.
82. Belokon YN. Chiral complexes of Ni (II) Cu (II), and Cu (I) as reagents, catalysts and receptors for asymmetric synthesis and chiral recognition of amino acids. *Pure and applied chemistry*. 1992. **64**(12): 1917–24.
83. Ueki H., Ellis T. K., Martin C. H. *et al.* Improved Synthesis of Proline Derived Ni(II)-Complexes of Glycine, a Versatile Chiral Equivalents of Nucleophilic Glycine for General Asymmetric Synthesis of α -Amino Acids. *J. Org. Chem.* 2003. **68**: 7104–7107.
- DOI: 10.1021/jo0301494.
84. Soloshonok V. A., Avilov D. V., Kukhar V. P., *et al.* An Efficient Asymmetric Synthesis of (2S, 3S)-3-Trifluoromethylpyroglutamic Acid. *Tetrahedron Lett.* 1997. **38**: 4903–4904.
85. Soloshonok V. A., Cai C., Hruby V. J. *et al.* Stereochemically Defined C-Substituted Glutamic Acids and their Derivatives. 1. An Efficient Asymmetric Synthesis of (2S, 3S)-3-Methyl- and -3-Trifluoromethylpyroglutamic Acids. *Tetrahedron* 1999. **55**: 12031–12044.
86. Soloshonok V. A., Kirilenko A. G., Kukhar V. P., Resnati G. Transamination of Fluorinated β -Keto Carboxylic Esters. A Biomimetic Approach to β -Polyfluoroalkyl- β -Amino Acids, *Tetrahedron Lett.* 1993. **34**: 3621–3624.
87. Soloshonok V. A., Kirilenko A. G., Galushko S. V., Kukhar V. P. Catalytic Asymmetric Synthesis of β -Fluoroalkyl- β -Amino Acids via Biomimetic [1,3]-Proton Shift Reaction. *Tetrahedron Lett.* 1994. **35**: 5063–5064.
88. Bravo P., Capelli S., Meille S. V. *et al.* Synthesis of Optically Pure (R)- and (S)- α -Trifluoromethyl-Alanine. *Tetrahedron: Asymmetry* 1994. **5**: 2009–2018.
89. Soloshonok V. A., Cai C., Hruby V. J., Meervelt L. V. Asymmetric Synthesis of Novel Highly Sterically Constrained (2S, 3S)-3-Methyl-3-Trifluoromethyl- and (2S, 3S, 4R)-3-Trifluoromethyl-4-Methylpyroglutamic Acids, *Tetrahedron*. 1999. **55**: 12045–12058.
90. Soloshonok V. A., Kukhar V. P., Galushko S. V., *et al.* General method for the synthesis of enantiomerically pure β -hydroxy- α -amino acids, containing fluorine atoms in the side chains. Case of stereochemical distinction between methyl and trifluoromethyl groups. X-ray crystal and molecular structure of the Nickel(II) complex of (2S,3S)-2-(trifluoromethyl) threonine. *J. Chem. Soc., Perkin Trans. I*. 1993. 3143–3155.
91. Soloshonok V. A., Avilov D. V., Kukhar V. P. Highly Diastereoselective Asymmetric Aldol Reactions of Chiral Ni(II)-Complex of Glycine with Trifluoromethyl Ketones. *Tetrahedron: Asymmetry*. 1996. **7**: 1547–1550.

92. Cai C., Soloshonok V. A., Hraby V. J. Michael Addition Reactions Between Chiral Ni(II) Complex of Glycine and 3-(trans-enoyl)oxazolidin-2-ones. A Case of Electron Donor-Acceptor Attractive Interactions-Controlled Face Diastereoselectivity. *J. Org. Chem.* 2001. **66**: 1339–1350.
93. Soloshonok V. A., Cai C., Hraby V. J. Asymmetric Michael Addition Reactions of Chiral Ni(II) Complex of Glycine with N-(Enoyl)oxazolidinones: Improved Reactivity and Stereochemical Outcome. *Tetrahedron: Asymmetry*. 1999. **10**: 4265–4269.
94. Soloshonok V. A., Cai C., Hraby V. J. Toward Design of a Practical Methodology for Stereocontrolled Synthesis of χ -Constrained Pyroglutamic Acids and Related Compounds. Virtually Complete Control of Simple Diastereoselectivity in the Michael Addition Reactions of Glycine Ni(II) Complexes with N-(Enoyl)oxazolidinones. *Tetrahedron Lett.* 2000. **41**: 135–139.
95. Soloshonok V. A., Hayashi T., Ishikawa K., Nagashima N. Highly Diastereoselective Aldol Reaction of Fluoroalkyl Aryl Ketones with Methyl Isocyanoacetate Catalyzed by Silver(I)/Triethylamine. *Tetrahedron Lett.* 1994. **35**: 1055–1058.
96. Soloshonok V. A., Hayashi T. Gold(I)-Catalyzed Asymmetric Aldol Reaction of Fluorinated Benzaldehydes with α -Isocyanoacetamide. *Tetrahedron: Asymmetry* 1994. **5**: 1091–1094.
97. Soloshonok V. A., Kacharov A. D., Hayashi T. Gold(I)-Catalyzed Asymmetric Aldol Reactions of Isocyanoacetic Acid Derivatives with Fluoroaryl Aldehydes. *Tetrahedron*. 1996. **52**: 245–254.
98. Evans D.A., Chapman K.T., Bisaha J. Asymmetric Diels-Alder cycloaddition reactions with chiral. α ., β -unsaturated N-acyloxazolidinones. *J. Am. Chem. Soc.* 1988. **110**(4):1238–56.
99. Bordwell F.G. Equilibrium acidities in dimethyl sulfoxide solution. *Acc. Chem. Res.* 1988. **21**(12): 456–63.
100. Evans D.A., Anderson J.C., Taylor M.K. Studies directed toward the design of chiral acylating agents. The utility of chiral N-benzoylimides in enantioselective alcohol acylation. *Tetrahedron letters*. 1993. **34**: 5563–6.
101. Soloshonok V. A., Cai C., Hraby V. J. A Practical Asymmetric Synthesis of Enantiomerically Pure 3-Substituted Pyroglutamic Acids and Related Compounds. *Angew. Chem., Int. Ed.* 2000. **39**: 2172–2175.
102. Soloshonok V. A., Cai C., Yamada T., *et al.* Michael Addition Reactions Between Chiral Equivalents of a Nucleophilic Glycine and (S)- or (R)-3-(E-Enoyl)-4-phenyl-1,3-oxazolidin-2-ones as a General Method for Efficient Preparation of β -Substituted Pyroglutamic Acids. Case of Topographically Controlled Stereoselectivity. *J. Am. Chem. Soc.* 2005. **127**: 15296–15303.
103. Soloshonok V. A., Cai C., Hraby V. J. *et al.* Rational Design of Highly Diastereoselective, Organic Base-Catalyzed, Room Temperature Michael Addition Reactions, *J. Org. Chem.* 2000. **65**: 6688–6696.
104. Soloshonok V. A., Ueki H., Tiwari R., *et al.* Virtually Complete Control of Simple and Face Diastereoselectivity in the Michael Addition Reactions between Achiral Equivalents of a Nucleophilic Glycine and (S)- or (R)-3-(E-Enoyl)-4-phenyl-1,3-oxazolidin-2-ones: Practical Method for Preparation of β -Substituted Pyroglutamic Acids and Prolines. *J. Org. Chem.* 2004. **69**: 4984–4990.
105. Soloshonok V. A., Cai C., Hraby V. J. A Unique Case of Face Diastereoselectivity in the Michael Addition Reactions Between Ni(II)-Complexes of Glycine and Chiral 3-(E-enoyl)-1,3-oxazolidin-2-ones. *Tetrahedron Lett.* 2000. **41**: 9645–9649.
106. Soloshonok V. A., Cai C., Hraby V. J. (S)- or (R)-N-(E-enoyl)-4-phenyl-1,3-oxazolidin-2-ones: Ideal Michael Acceptors to Afford a Virtually Complete Control of Simple and Face Diastereoselectivity in Addition Reactions with Glycine Derivatives. *Org. Lett.* 2000. **2**: 747–750.
107. Ueki H., Ellis T. K., Martin C. H., Soloshonok V. A. Efficient Large-Scale Synthesis of Pico-

- linic Acid Derived Ni(II)-Complexes of Glycine. *Eur. J. Org. Chem.* 2003. 1954–1957. DOI: 10.1002/ejoc.200200688
108. Soloshonok V. A., Ueki H., Ellis T. K. *et al.* Application of Modular Nucleophilic Glycine Equivalents for Truly Practical Asymmetric Synthesis of β -Substituted Pyroglutamic Acids. *Tetrahedron Lett.* 2005. **46**: 1107–1110.
109. Ellis T. K., Ueki H., Yamada T. *et al.* The Design, Synthesis and Evaluation of a New Generation of Modular Nucleophilic Glycine Equivalents for the Efficient Synthesis of Sterically Constrained α -Amino Acids. *J. Org. Chem.* 2006. **71**: 8572–8578.
110. Zou Y., Han J., Saghyan A. S. *et al.* Asymmetric Synthesis of Tailor-Made Amino Acids Using Chiral Ni(II)-Complexes of Schiff Bases; An Update of the Recent Literature. *Molecules*. 2020. **25**: 2739. doi:10.3390/molecules25122739.
111. Soloshonok V. A., Ueki H., Ellis T. K. New Generation of Nucleophilic Glycine Equivalents, *Tetrahedron Lett.* 2005. **46**: 941–944.
112. Romoff T. T., Palmer A. B., Mansour N. *et al.* Scale-up Synthesis of (R)- and (S)-N-(2-Benzoyl-4-chlorophenyl)-1-(3,4-dichlorobenzyl) pyrrolidine-2-carboxamide Hydrochloride, a Versatile Reagent for Preparation of Tailor-made α - and β -Amino Acids in Enantiomerically Pure Form. *Org. Process Res. Dev.* 2017. **21**: 732–739. DOI: 10.1021/acs.oprd.7b00055.
113. Kawamura A., Moriwaki H., Röschenhaler G.-V., *et al.* Synthesis of (2S,3S)- β -(trifluoromethyl)- α,β -diamino acid by Mannich addition of glycine Schiff base Ni(II) complexes to N-tert-butylsulfinyl-3,3,3-trifluoroacetaldehyde. *J. Fluor. Chem.* 2015. **171**: 67–72. DOI: 10.1016/j.jfluchem.2014.09.013.
114. Li T., Zhou S., Wang J., *et al.* Asymmetric synthesis of α -(1-oxoisindolin-3-yl)glycine: synthetic and mechanistic challenges. *Chem. Commun.* 2015. **51**: 1624–1626. DOI: 10.1039/C4CC05659K.
115. Nian Y., Wang J., Moriwaki H. *et al.* Analysis of crystallographic structures of Ni(II) complexes of α -amino acid Schiff bases; Elucidation of the substituents effect on stereochemical preferences. *Dalton Trans.* 2017. **46**: 4191–4198. DOI: 10.1039/C7DT00014F.
116. Stierhof M., Hansen K. Ø., Sharma M. *et al.* New cytotoxic callipeltins from the Solomon Island marine sponge *Asteropus* sp. *Tetrahedron*. 2016. **72**: 6929–6934.
117. Tokairin Y., Soloshonok V. A., Moriwaki H., Konno H. Asymmetric synthesis of (2S,3S)-3-Me-glutamine and (R)-allo-threonine derivatives proper for solid phase peptide coupling. *Amino Acids*. 2019. **51**: 419–432. https://doi.org/10.1007/s00726-018-2677-5.
118. Cheng L., Goodwin C. A., Schully M. F. *et al.* Synthesis and biological activity of ketomethylene pseudopeptide analogs as thrombin inhibitors. *J. Med. Chem.* 1992. **35**: 3364–3369.
119. Pavar M. C., Hanif K., Azam A. *et al.* Structure–activity relationship study between ornithyl-proline and lysyl-proline based tripeptidomimics as angiotensin converting enzyme inhibitors. *Bioorg. Med. Chem. Lett.* 2006. **16**: 2117–2121.
120. Stranix B. R., Lavallée J.-F., Sévigny G., *et al.* Lysine sulfonamides as novel HIV-protease inhibitors: N ϵ -acyl aromatic α -amino acids. *Bioorg. Med. Chem. Lett.* 2006. **16**: 3459–3462.
121. Brust A., Palant E., Croker D. E., *et al.* x-Conopeptide pharmacophore development: toward a novel class of norepinephrine transporter inhibitor (Xen2174) for pain. *J. Med. Chem.* 2009. **52**: 6991–7002.
122. Lee Y. S., Nyberg J., Moye S., *et al.* Understanding the structural requirements of 4-anilidopiperidine analogues for biological activities at m and s opioid receptors. *Bioorg. Med. Chem. Lett.* 2007. **17**: 2161–2165.
123. Zhu Y., Zhang W., Mei H., *et al.* Catalytic enantioselective Michael addition reactions of tertiary enolates generated by detrifluoroacetylation. *Chem.–Eur. J.* 2017. **23**: 11221–11225. DOI: 10.1002/chem.201702091.
124. Nagaoka K., Mei H., Guo Y., *et al.* Michael addition reactions of chiral glycine Schiff base Ni

- (II)-complex with 1-(1-phenylsulfonyl)benzene. *Chirality* 2020. **32**: 885–893.
<https://doi.org/10.1002/chir.23203>.
125. Shigeno Y., Han J., Soloshonok V. A., *et al.* Asymmetric synthesis of (S)-3-methylene-glutamic acid and its N-Fmoc derivative via Michael addition of chiral glycine Schiff base Ni(II) complex with enol tosylates. *Chirality* 2021. **33**: 115–123.
126. Yamada T., Okada T., Sakaguchi K., *et al.* Efficient Asymmetric Synthesis of Novel 4-Substituted and Configurationally Stable Analogs of Thalidomide. *Org. Lett.* 2006. **8**: 5625–5628.
127. Soloshonok V. A., Yamada T., Sakaguchi K., Ohfuné Y. Concise Asymmetric Synthesis of Configurationally Stable 4-trifluoromethyl Thalidomide. *Future Med. Chem.* 2009. **1**: 897–908. doi.org/10.4155/fmc.09.63.
128. King F. E., Kidd D. A. Novel Method for the Synthesis of γ -L-Glutamyl Dipeptides Based on a Complexing Co-protection Strategy. *J. Org. Chem.* 1949. **14**: 3315–3321.
129. Flaih N., Pham-Huy C., Galons H. An expeditious synthesis of cyclic imides. *Tetrahedron Lett.* 1999. **40**: 3697–38369.
130. Yamada T., Sakaguchi K., Shinada T., *et al.* Efficient asymmetric synthesis of the functionalized pyroglutamate core unit common to oxazolomycin and neooxazolomycin using Michael reaction of nucleophilic glycine Schiff base with α,β -disubstituted acrylate. *Tetrahedron: Asymmetry*. 2008. **19**: 2789–2795. doi.org/10.1016/S0040-4020(98)00779-0.
131. Jörres M., Aceña J. L., Soloshonok V. A., Bolm C. Asymmetric Carbon-Carbon Bond Formations under Solvent-Less Conditions in Ball Mills, *Chem. Cat. Chem.* 2015. **7**: 1265–1269; DOI: 10.1002/cctc.201500102.
132. Bergagnini M., Fukushi K., Han J. *et al.* NH-type of chiral Ni(II) complexes of glycine Schiff base: design, structural evaluation, reactivity and synthetic applications. *Org. Biomol. Chem.* 2014. **12**: 1278–1291. DOI: 10.1039/C3OB41959B.
133. Wang J., Lin D., Zhou S. *et al.* Asymmetric Synthesis of Sterically Constrained Linear Trifluoromethyl Containing Amino Acids via Alkylation of Chiral Equivalents of Nucleophilic Glycine and Alanine. *J. Org. Chem.* 2011. **76**: 684–687.
134. Soloshonok V. A., Avilov D. V., Kukhar V. P. Asymmetric Aldol Reactions of Trifluoromethyl Ketones with a Chiral Ni(II) Complex of Glycine: Stereocontrolling Effect of the Trifluoromethyl Group. *Tetrahedron*. 1996. **52**: 12433–12442.
135. Soloshonok V. A., Avilov D. V., Kukhar V. P. Asymmetric Aldol Reactions of Chiral Ni(II)-Complex of Glycine with Aldehydes. Stereodivergent Synthesis of syn-(2S)- and syn-(2R)- β -Alkylserines, *Tetrahedron: Asymmetry* 1995. **6**: 1741–1756.
136. Jörres M., Chen X., Aceña J. L. *et al.* Asymmetric Synthesis of α -Amino Acids under Operationally Convenient Conditions, *Adv. Synth. Catal.* 2014. **356**: 2203–2208. DOI: 10.1002/adsc.201400405.
137. Soloshonok V. A., Avilov D. V., Kukhar V. P., *et al.* Highly Diastereoselective aza-Aldol Reactions of a Chiral Ni(II) Complex of Glycine with Imines. An Efficient Asymmetric Approach to 3-Perfluoroalkyl-2,3-Diamino Acids. *Tetrahedron Lett.* 1997. **38**: 4671–4674.
138. Sorochinsky A. E., Ueki H., Aceña J. L. *et al.* Chemical approach for interconversion of (S)- and (R)- α -amino acids. *Org. Biomol. Chem.* 2013. **11**: 4503–4507. DOI:10.1039/C3OB40541A.
139. Zhou S., Wang J., Chen X. *et al.* Chemical Kinetic Resolution of Unprotected β -Substituted- β -Amino Acids Using Recyclable Chiral Ligands. *Angew. Chem. Int. Ed.* 2014. **53**: 7883–7886. DOI: 10.1002/anie.201403556.

Стаття надійшла 25.05.2024.